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# FLUORESCENCE DECAY STUDIES OF POLY(RIBOADENYLIC ACID) CONTAINING 1-N-6-ETHENOADEN-OSINE

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The fluorescence properties of the  $1-N^6$ -etheno derivatives of poly(riboadenylic acid) (poly(rA, $\epsilon$ rA)) have been examined. The fluorescence quantum yield of poly(rA, $\epsilon$ rA) decreases with an increase in the degree of the  $\epsilon$ A substitution and is much smaller than that for  $\epsilon$ AMP even for low degrees of  $\epsilon$ A substitution. The nearest-neighbor interactions such as  $\epsilon$ -adenine-adenine and  $\epsilon$ -adenine may be responsible for this behavior. It is found that the fluorescence decay kinetics obeys a three-exponential decay law for poly(rA, $\epsilon$ rA), suggesting that there exist at least three different stacked conformational states.

### 1. Introduction

Because of the stability, high fluorescence yields and relatively long fluorescence lifetimes of  $1-N^6$ ethenoadenosine ( $\epsilon A$ ) and its nucleotides [1-3]. they may serve as excellent probes for studying the conformation of nucleic acids. It is well established that the conformation of poly(riboadenylic acid) (poly(rA)) in solution changes from the hydrogen-bonded double-strand helix at low pH to the ordered single-strand structure of stacked bases at neutral pH [4]. Therefore, it is of great interest to study the fluorescence behavior of poly(rA) containing  $\epsilon A$  groups (poly(rA, $\epsilon rA$ )) in connection with its conformational change. Steiner and coworkers [5] first investigated poly(rA, erA) using fluorescence polarization techniques and showed that the intensity and polarization of fluorescence respond to helix-coil transitions.

In the preceding paper [6], we have reported that the fluorescence decay kinetics of dinucleoside monophosphates containing  $\epsilon A$ , which depends on both the kind of bases and the base sequence, obeys a three-exponential decay law; it

has been proposed that there are three or more conformational states in  $\epsilon$ -dinucleoside monophosphates. To elucidate further the base-stacking interactions in polynucleotides, we have reexamined the fluorescence of poly(rA, $\epsilon$ rA) by steady-state and transient decay measurements. It is found in the present study that the fluorescence decay kinetics of poly(rA, $\epsilon$ rA) also follows a three-exponential decay law at both acid (pH 5) and neutral pH (pH 7), suggesting that there exist at least three different stacked conformational states in poly(rA, $\epsilon$ rA). Preliminary results have been recently reported [7].

#### 2. Materials and methods

Poly(rA) was purchased from Seikagaku Kogyo and was used without further purification ( $s_{20,w}^0 = 10.7$ ).  $\epsilon$ A groups were introduced into poly(rA) according to the procedure described by Steiner et al. [5]. The degree of the  $\epsilon$ A substitution was controlled by varying the reaction time. After completion of the reaction, the pH of the solution

was adjusted to about 7, and then the solution was dialyzed against the desired buffer solution at  $4^{\circ}$ C. Approximate values of  $\epsilon$ AMP contents of the derivatives were determined from the absorption spectrum according to the method of Steiner et al. [5]. Glass-redistilled water and analytical grade reagents were used for the preparation of all solutions.

Absorption and steady-state fluorescence spectra were measured with a Shimadzu UV-200S spectrophotometer and a Hitachi MPF-2A spectrophotofluorometer, respectively. Observed fluorescence spectra were corrected for the unequal quantum response of the detector system which consists of lenses, a monochromator and an R106-UH photomultiplier tube (Hamamatsu TV). Fluorescence quantum yields of the  $\epsilon$ A derivatives were determined by comparing the area under the fluorescence spectrum with the corresponding area of  $\epsilon$ AMP and by taking 0.52 for the quantum yield of  $\epsilon$ AMP [8]. Transient fluorescence decay measurements and the analysis of data were described in the preceding paper [6].

All measurements were carried out using two solvent systems: 0.005 M sodium phosphate, 0.1 M NaCl (pH 7.0) and 0.01 M sodium acetate, 0.1 M NaCl (pH 5.0). The concentrations of poly(rA, $\epsilon$ rA) were in the range  $5.0 \times 10^{-5}$ – $10^{-4}$  M in monomer units.

# 3. Results

# 3.1. Absorption and fluorescence spectra

Fig. 1 shows absorption spectra for poly(rA) and poly(rA, $\epsilon$ rA) at pH 7. The absorption spectra of poly(rA, $\epsilon$ rA) resemble qualitatively the expected summation of AMP and  $\epsilon$ AMP contributions.

Fig. 2 summarizes the temperature dependence of absorbance at around 260 nm at pH 7. Poly(τA,ετA) derivatives show a decrease in hypochroism with increasing temperature. This behavior is similar to the thermally induced spectral change observed for the single-stranded helical form of poly(τA) itself [4]. In parallel to changes observed for poly(τA), the increase in absorbance

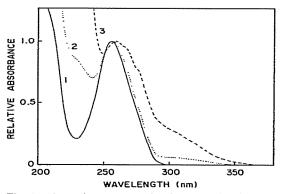


Fig. 1. Absorption spectra of poly(rA) and poly(rA, εrA) in 0.005 M sodium phosphate. 0.1 M NaCl, pH 7.0. 25°C: (1) poly(rA). (2) poly(rA<sub>0.65</sub>, εrA<sub>0.35</sub>). (3) poly(rA<sub>0.20</sub>, εrA<sub>0.80</sub>). The maximum of each spectrum (around 260 nm) is normalized to unity.

is gradual and shows no qualitative indication of any cooperative sharpening. It should be noted that the hyperchromicity decreases with increasing degree of  $\epsilon A$  substitution (fig. 2). In contrast to the results at pH 7, the increase in absorbance at pH 5 indicates a cooperative sharpening (fig. 3). This behavior responds to helix-coil transitions [4]. As seen in fig. 3, both the transition tempera-

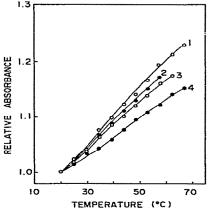


Fig. 2. Temperature dependence of absorbance for poly(rA) and poly(rA,  $\epsilon$ rA) in 0.005 M sodium phosphate, 0.1 M NaCl, pH 7.0: (1) poly(rA) ( $\lambda$  = 256 nm), (2) poly(rA<sub>0.90</sub>,  $\epsilon$ rA<sub>0.10</sub>) ( $\lambda$  = 256 nm), (3) poly(rA<sub>0.65</sub>,  $\epsilon$ rA<sub>0.35</sub>) ( $\lambda$  = 257 nm), (4) poly(rA<sub>0.20</sub>,  $\epsilon$ rA<sub>0.80</sub>) ( $\lambda$  = 258 nm).

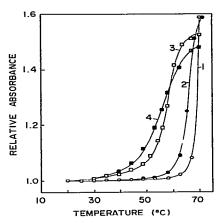


Fig. 3. Temperature dependence of absorbance for poly(rA) and poly(rA,  $\epsilon$ rA) in 0.01 M sodium acetate, 0.1 M NaCl, pH 5.0: (1) pciy(rA) ( $\lambda$  = 251 nm), (2) poly(rA<sub>0.97</sub>,  $\epsilon$ rA<sub>0.03</sub>) ( $\lambda$  = 251 nm), (3) poly(rA<sub>0.90</sub>,  $\epsilon$ rA<sub>0.10</sub>) ( $\lambda$  = 252 nm), (4) poly(rA<sub>0.65</sub>,  $\epsilon$ rA<sub>0.35</sub>) ( $\lambda$  = 252 nm).

ture and the hyperchromic change decrease with an increase in degree of  $\epsilon A$  substitution. In view of these facts, it is concluded that the introduction of  $\epsilon A$  may destabilize the ordered structure of poly(rA).

As depicted in fig. 4, the fluorescence spectrum of poly(rA, $\epsilon$ rA) at both pH 5 and pH 7 shows a wavelength dependence similar to that of  $\epsilon$ AMP

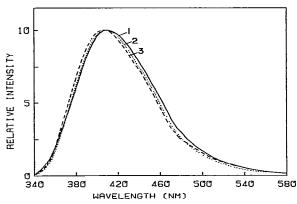


Fig. 4. Normalized fluorescence quantum spectra of  $\epsilon$ AMP and poly(rA, $\epsilon$ rA) at 25°C: (1)  $\epsilon$ AMP (pH 5.0, pH 7.0), (2) poly(rA<sub>0.90</sub>, $\epsilon$ rA<sub>0.10</sub>) (pH 7.0), (3) poly(rA<sub>0.90</sub>, $\epsilon$ rA<sub>0.10</sub>) (pH 5.0).

except that its maximum wavelength for low degrees of  $\epsilon A$  substitution (3-35%) slightly shifts to shorter wavelength. This means that  $\epsilon A$  groups are embedded into the hydrophobic region. The magnitude of the spectral shift was greater at pH 5 than at pH 7, suggesting that the microenvironment of  $\epsilon A$  is more hydrophobic at pH 5. On the other hand, the fluorescence spectrum for a high degree of  $\epsilon A$  substitution (80%) was very similar to that of  $\epsilon AMP$  itself, indicating that  $\epsilon A$  groups are considerably exposed to the surrounding solvent.

#### 3.2. Fluorescence quantum yields

The fluorescence quantum yields  $(\Phi_F)$  for poly(rA,  $\epsilon$ rA) markedly decreased with increasing degree of  $\epsilon$ A substitution (fig. 5). The  $\Phi_F$  values for poly(rA,  $\epsilon$ rA) even for low degrees of substitution are much smaller than that for  $\epsilon$ AMP (fig. 5, and tables 1 and 2). In the preceding paper [6], it was also found that the  $\Phi_F$  value for  $\epsilon$ AP $\epsilon$ A is much smaller than that for  $\epsilon$ AMP. In view of the fact that adenine residues can quench the fluorescence of  $\epsilon$ AMP [8], the nearest-neighbor interactions between  $\epsilon$ -adenine and adenine or  $\epsilon$ -adenine and  $\epsilon$ -adenine may be responsible for fluorescence

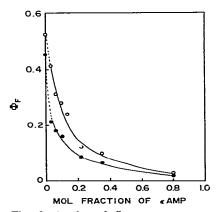


Fig. 5. A plot of fluorescence quantum yields  $(\Phi_F)$  for poly(rA,  $\epsilon$ rA) against the degree of  $\epsilon$ A substitution: ( $\bigcirc$ ) 0.005 M sodium phosphate, 0.1 M NaCl, pH 7.0, 25°C; ( $\bullet$ ) 0.01 M sodium acetate, 0.1 M NaCl, pH 5.0, 25°C. The  $\Phi_F$  values where the mole fraction of  $\epsilon$ AMP is zero are those for  $\epsilon$ AMP itself.

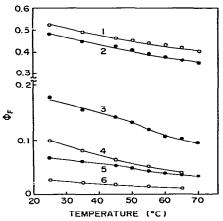


Fig. 6. Temperature dependence of fluorescence quantum yields  $(\Phi_F)$  for  $\epsilon$ AMP and poly(rA, $\epsilon$ rA): (1)  $\epsilon$ AMP (pH 7.0), (2)  $\epsilon$ AMP (pH 5.0), (3) poly(rA<sub>0.90</sub>, $\epsilon$ rA<sub>0.10</sub>) (pH 5.0), (4) poly(rA<sub>0.65</sub>, $\epsilon$ rA<sub>0.35</sub>) (pH 7.0), (5) poly(rA<sub>0.65</sub>, $\epsilon$ rA<sub>0.35</sub>) (pH 5.0), (6) poly(rA<sub>0.20</sub>, $\epsilon$ rA<sub>0.80</sub>) (pH 7.0).

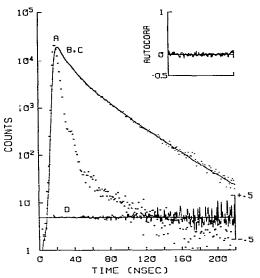


Fig. 7. Three-component analysis of the fluorescence decay of poly(rA<sub>0.65</sub>,  $\epsilon$ rA<sub>0.35</sub>) in 0.005 M sodium phosphate, 0.1 M NaCl, pH 7.0, 25°C. Curve A is the apparatus response function. Curve B is the observed decay curve. The smooth curve C shows the theoretical decay curve. Curve D is the weighted residuals. The inset is the autocorrelation function of the residuals. Parameters obtained:  $\tau_1 = 32.8$  ns,  $\tau_2 = 11.6$  ns,  $\tau_3 = 1.8$  ns,  $\alpha_1 = 0.047$ ,  $\alpha_2 = 0.093$ ,  $\alpha_3 = 0.213$  and  $\chi^2 = 1.29$ . The amplitudes normalized to unity are  $\alpha_1 = 0.13$ ,  $\alpha_2 = 0.27$  and  $\alpha_3 = 0.60$ .

quenching. A marked reduction in  $\Phi_F$  observed for highly substituted etheno derivatives poly( $rA_{0.20}$ ,  $\epsilon rA_{0.80}$ )) or  $\epsilon Ap\epsilon A$  suggests that homologous interactions of  $\epsilon$ -adenine are more efficient in producing quenching than interactions between  $\epsilon$ -adenine and adenine.

In analogy with the change in absorbance. the fluorescence quantum yield was dependent upon temperature (fig. 6).  $\epsilon$ AMP itself showed no temperature dependence of absorbance, while it was found to show an important degree of quenching with increasing temperature. This may arise from the increased rates of the radiationless deactivation processes which compete with direct emission. The  $\Phi_F$  value for poly(rA, $\epsilon$ rA) at pH 7 is gradually decreased with a rise in temperature, whereas the  $\Phi_F$  value at pH 5 shows a cooperative abrupt decrease at the transition temperature as well as the absorption change (cf. fig. 6 and figs. 2 and 3). In the present case, however, it should be noted that the effects of thermal destruction of the base

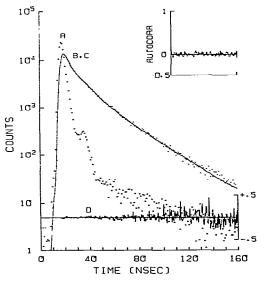


Fig. 8. Three-component analysis of the fluorescence decay of poly(rA<sub>0.65</sub>,  $\epsilon$ rA<sub>0.35</sub>) in 0.01 M sodium acetate, 0.1 M NaCl, pH 5.0, 25°C. Legends to each curve are as described for fig. 7. The theoretical decay curve is based on the following parameters:  $\tau_1 = 25.2$  ns,  $\tau_2 = 10.5$  ns,  $\tau_3 = 1.9$  ns,  $\alpha_1 = 0.032$ ,  $\alpha_2 = 0.077$ ,  $\alpha_3 = 0.203$  and  $\chi^2 = 1.38$ . The amplitudes normalized to unity are  $\alpha_1 = 0.10$ ,  $\alpha_2 = 0.25$  and  $\alpha_30.65$ .

Table 1 Fluorescence decay parameters and quantum yields for poly(rA,  $\epsilon$ rA) derivatives in 0.005 M sodium phosphate, 0.1 M NaCl, pH 7.0 The excitation and emission wavelengths were 337 and 410 nm, respectively; the results obtained here are not dependent on the emission wavelength (cf. table 3).  $\chi^2$  values ranged from 1.2 to 1.9. The amplitudes ( $\alpha$ ) are normalized to unity.

Preparation	Temperature (°C)	τ <sub>1</sub> (πs)	αι	τ <sub>2</sub> (ns)	$\alpha_2$	τ <sub>3</sub> (ns)	α3	$\Phi_{\mathrm{F}}$
€AMP	25	25.0	1.00	-				0.52
	45	22.6	1.00					0.46
	65	20.3	1.00					0.41
Poly(rA <sub>0.97</sub> , erA <sub>0.03</sub> )	25	36.3	0.65	16.1	0.15	2.4	0.20	0.41
	45	29.9	0.42	12.8	0.24	2.0	0.34	0.27
	65	21.0	0.33	9.2	0.25	1.8	0.42	0.17
$Poly(rA_{0.94}, \epsilon rA_{0.06})$	25	35.7	0.47	14.6	0.20	2.2	0.33	0.31
$Poly(rA_{0.90}, \epsilon rA_{0.10})$	25	35.4	0.44	13.8	0.24	2.2	0.32	0.28
$Poly(rA_{0.87}, \epsilon rA_{0.13})$	25	34.4	0.38	12.4	0.24	1.8	0.38	0.22
$Poly(rA_{0.78}, \epsilon rA_{0.22})$	25	33.7	0.21	12.1	0.32	2.0	0.47	0.12
$Poly(rA_{0.65}, \epsilon rA_{0.35})$	25	32.8	0.13	11.6	0.27	1.8	0.60	0.099
	45	27.3	0.08	8.5	0.28	1.4	0.64	0.064
	65	19.1	0.06	6.1	0.21	1.2	0.73	0.041
$Poly(rA_{0.20}, \epsilon rA_{0.80})$	25	26.9	0.05	8.6	0.20	1.4	0.75	0.026
	45	21.0	0.02	6.5	0.20	1.3	0.78	0.018
	65	21.9	0.003	5.1	0.16	1.0	0.84	0.011

Table 2
Fluorescence decay parameters and quantum yields for poly(rA,  $\epsilon$ rA) derivatives in 0.01 M sodium acetate, 0.1 M NaCl, pH 5.0
The excitation and emission wavelengths were 337 and 410 nm, respectively; the results obtained here are not dependent on the emission wavelength (cf. table 3).  $\chi^2$  values ranged from 1.1 to 1.9. The amplitudes ( $\alpha$ ) are normalized to unity.

Preparation	Temperature (°C)	τ <sub>ι</sub> (ns)	$\alpha_1$	τ <sub>2</sub> (ns)	$\alpha_2$	τ <sub>3</sub> (ns)	$\alpha_3$	$\Phi_{F}$
€AMP	25	20.7	1.00					0.45
	45	18.1	1.00					0.42
	65	16.5	1.00					0.36
$Poly(rA_{0.97}, \epsilon rA_{0.03})$	25	27.5	0.24	11.2	0.27	2.3	0.49	0.21
	45	24.6	0.17	10.5	0.29	1.8	0.54	0.18
	65	19.8	0.15	9.2	0.33	1.9	0.52	0.13
Poly(rA <sub>0.94</sub> , erA <sub>0.06</sub> )	25	28.3	0.23	12.0	0.33	2.8	0.44	0.18
$Poly(rA_{0.90}, \epsilon rA_{0.10})$	25	28.2	0.24	12.2	0.29	2.5	0.47	0.16
	45	25.3	0.13	11.0	0.30	2.1	0.57	0.14
	65	23.6	0.06	10.3	0.35	2.2	0.59	0.10
$Poly(rA_{0.78}, \epsilon rA_{0.22})$	25	26.6	0.13	11.3	0.27	2.1	0.60	0.084
$\operatorname{Poly}(\operatorname{rA}_{0.65},\epsilon\operatorname{rA}_{0.35})$	25	2.5.2	0.10	10.5	0.25	1.9	0.65	0.067
	45	22.6	0.06	8.9	0.25	1.7	0.69	0.052
	65	19.8	0.03	6.4	0.21	1.1	0.76	0.037
$Poly(rA_{0.20}, \epsilon rA_{0.80})$	25	20.5	0.04	7.4	0.14	1.1	0.82	0.024

stacking are superimposed upon those of enhanced thermal quenching of the excited state.

### 3.3. Fluorescence decay curves

It was found that the fluorescence decay kinetics of  $\epsilon$ AMP followed a single-exponential decay law at both pH 5 and pH 7, while the decay behavior for poly( $rA, \epsilon rA$ ), which depends upon temperature and pH, was rather complex. Typical decay curves obtained with poly(rA<sub>0.65</sub>, \( \epsilon rA<sub>0.35</sub> ) are shown in figs. 7 and 8. All decay curves clearly indicated the deviation from the single exponentiality. The reduced  $\chi^2$ , the weighed residuals and the autocorrelation function of the residuals indicated that the decay data could not be explained in terms of a double-exponential decay law but could well be resolved into three-exponential components corresponding to the long  $(\tau_1)$ , medium  $(\tau_2)$  and short  $(\tau_3)$  lifetimes. Typical sets of decay parameters together with  $\Phi_{\mathrm{F}}$  are summarized in tables 1 and 2. The  $\chi^2$  values obtained were in the range 1.1-1.9.

Time-resolved fluorescence spectra on the nanosecond scale were measured in order to study any time-dependent interactions. The spectra were taken at time intervals ranging from 0 to 150 ns after the peak of the exciting pulse. It was found that spectra showed no time dependence and were nearly identically with the steady-state fluorescence spectra. In harmony with this finding, the

decay parameters did not show any significant dependence on the emission wavelength (table 3).

#### 4. Discussion

It is found in the present study that the fluorescence decay kinetics of  $poly(rA, \epsilon rA)$  follows a three-exponential decay law. As is the case with dinucleoside monophosphates containing  $\epsilon A$  [6], two explanations for this behavior are possible: one is a reversible excited-state reaction and the other is the existence of three different conformational states.

Considerable stacking interactions between two bases are necessary to produce efficient fluorescence quenching. Relaxation kinetic studies indicate that the stacking of bases in poly(rA) takes  $10^{-6}-10^{-7}$  s, and double-stranded helix formation in oligomers ranges from  $10^{-3}$  to  $10^{-1}$  s [9-11]. Taking into account the restrictions of sugar puckering and backbone rotations in polynucleotide chains, it seems unlikely that a reversible reaction like unstacked  $\rightleftharpoons$  stacked occurs during the lifetime of the excited state. In fact, the finding that nanosecond time-resolved fluorescence spectra of poly(rA, $\epsilon$ rA) are independent of time (cf. table 3) suggests that the environment of the  $\epsilon$ A group is not substantially altered during its lifetime.

We propose here another explanation which would assumed that the decay behavior has its

Table 3

Decay parameters obtained for poly(rA<sub>0.65</sub>,  $\epsilon$ rA<sub>0.35</sub>) at 25°C

The excitation wavelength was 337 nm.  $\chi^2$  values ranged from 1.2 to 1.8. The amplitudes ( $\alpha$ ) are normalized to unity.

Solvent	Emission wavelength (nm)	τ <sub>1</sub> (ns)	$\alpha_{\mathfrak{i}}$	τ <sub>2</sub> (ns)	$\alpha_2$	73 (ns)	$\alpha_3$
pH 7 4	410	32.8	0.13	11.6	0.27	1.8	0.60
	440	33.1	0.13	11.9	0.26	1.6	0.61
	470	31.9	0.14	11.5	0.27	1.5	0.59
	500	32.2	0.13	11.4	0.27	1.5	0.60
рН 5 <sup>ь</sup>	410	25.2	0.10	10.5	0.25	1.9	0.65
	440	25.7	0.10	10.3	0.26	1.9	0.64
	470	25.9	0.09	10.7	0.25	2.0	0.66

<sup>4 0.005</sup> M sodium phosphate, 0.1 M NaCl.

<sup>&</sup>lt;sup>b</sup> 0.01 M sodium acetate, 0.1 M NaCl.

origin in different conformational states, depending upon the stacking interaction between εadenine and its neighbors ( $\epsilon$ -adenine or adenine). The present decay data are not compatible with the two-state model proposed for a conformational change in dinucleoside phosphates [12,13]. The three-exponential decay kinetics for poly(rA.erA) suggests that the fluorescent states of  $\epsilon A$  groups consist of at least three different classes of conformations. Temperature-jump relaxation studies on single-stranded poly(rA) reveal that there are two relaxation times for its conformational change, demonstrating the existence of more than two independent conformational states [9,10]. Laser temperature-jump techniques also suggest that the relaxation kinetics of poly(rA), which depends upon ionic strength, is complicated and that there are more than two conformational states at high ionic strengths [14].

Now, the structure of these conformations disclosed by fluorescence decay measurements must be sought. It can be clearly seen in tables 1 and 2 that the amplitude  $\alpha_1$  decreases with increasing degree of  $\epsilon A$  substitution, whereas  $\alpha_3$  increases. It is proposed, therefore, that the long lifetime  $(\tau_1)$  is ascribed to unstacked  $\epsilon A$  groups, while the short lifetime  $(\tau_3)$  is a result of strong stacking interactions between  $\epsilon$ -adenine and its neighbors ( $\epsilon$ adenine or adenine). The medium lifetime  $(\tau_2)$  is presumably due to weak stacking interactions. The long lifetime  $(\tau_1)$  for low degrees of  $\epsilon A$  substitution at 25°C is 34-36 ns at pH 7 and 27-28 ns at pH 5, being much larger than the corresponding lifetimes for  $\epsilon$ AMP. This increase in lifetime may be due, in part, to the immersion of  $\epsilon A$  groups in the hydrophobic region where they are protected against quenching by collisions with the surrounding solvent. The  $\tau_1$  value decreases with an increase in degree of  $\epsilon A$  substitution and is very close to that for  $\epsilon$ AMP itself for the highest degree of substitution (poly( $rA_{0.20}$ ,  $\epsilon rA_{0.80}$ )) (tables 1 and 2). The  $\tau_1$  value at 65°C is also nearly equal to that of  $\epsilon$ AMP. It is expected that the ordered structure is considerably ruptured for a high degree of substitution or at high temperature. Under these conditions,  $\epsilon A$  groups are exposed to the surrounding solvent and hence, the lifetime becomes closer to that of  $\epsilon$ AMP. As seen in tables 1 and 2, the

amplitude  $\alpha_3$  a'  $\rho$ H 5 is larger than at pH 7. This demonstrates that there exist more efficient stacking interactions at pH 5 due to the ordered double-helical structure.

The three-exponential decay law is also found for  $\epsilon$ -dinucleoside phosphates [6]. However, the decay behavior of poly( $\tau A, \epsilon \tau A$ ) differs from that of  $\epsilon$ -dinucleoside phosphates. Especially, for low degrees of  $\epsilon A$  substitution, the amplitude  $\alpha_1$  and the lifetime  $\tau_1$  are considerable larger than those of  $\epsilon$ -dinucleoside phosphates (tables 1 and 2; and table 1 in ref. 6). Because of the limitations of sugar puckering and backbone rotations in polynucleotide chains, it seems likely that although  $\epsilon A$  groups in poly( $\tau A, \epsilon \tau A$ ) are immersed in the hydrophobic region, all of them cannot orient with their neighbors in favorable geometries to produce efficient quenching.

In conclusion, the present decay data indicate that  $poly(rA, \epsilon rA)$  can be described as consisting of at least three different conformational states. It may be that fluorescence decay measurements can be used to reveal the multistate behavior in other polynucleotide systems

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